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pluriBead®

Application Example

Monocyte isolation with pluriBead[®] for maturation of dendritic cells



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pluriSelect USA

Spring Valley, CA 91977 USA

Phone: 619-928-9265 support.usa@pluriselect.com sales.usa@pluriselect.com

www.pluriselect.com

pluriSelect Worldwide

Deutscher Platz 5c 04103 Leipzig Germany

Phone: +49 341 333858-0 support@pluriselect.com sales@pluriselect.com



Monocyte isolation with pluriBead[®] for maturation of dendritic cells

Sample volume	10 ml buffy coat
Isolation method	280 μl CD14 S-pluriBead® anti-Hu <i>or</i> 300 μl CD14 M-pluriBead® anti-Hu
Yield	~6 x 10 ⁶ monocytes with S-pluriBead® ~12 x 10 ⁶ monocytes with M-pluriBead®
Vitality	>92% (trypan blue staining)
Purity	~97%

Monocyte isolation, differentiation and stimulation protocol

To reduce soluble CD14 wash sample material twice with the provided Washing Buffer. Add CD14 pluriBead[®] for isolation of monocytes into the sample tube. Incubate on a pluriPlix[®] or rolling mixer for 20 minutes. Detach the CD14 cells from pluriBead[®].

Resuspend the cells in 1 ml of RPMI 1640 medium (plus 10% FCS and 1x Pen/Strep). Determine the cell number. Use 1 million cells per well for the cultivation of monocyte in a 24-well cell culture plate. Incubate with 1 ml monocyte culture medium (RPMI 1640 plus 10% FCS, 1x Pen/Strep, 2,000 U/ml GM-CSF and 200 U/ml IL-4) at 37°C and 5% carbon dioxide.

Remove the medium and add 1 ml fresh monocyte culture medium after 24 hours. Culture the cells for 4 more days. Stimulate the cells with 100 ng/ml LPS for another 24 hours (after a total of 5 days). Remove the activated cells with trypsin. Analyze the maturation rate to dendritic cells from monocytes by fluorescent analysis of CD1a+, CD14+ and CD83+.





CD14+ cells after isolation in culture

Maturation of monocytes to dendritic cells after 5 days of culture with GM-CSIF/IL-4 and stimulation with LPS

Fluorescent analysis of monocytes

